



Yield Decline and Resistance Development in Sucking Pests of Cotton in the Context of Unwise Spraying Techniques

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Abstract: Over 100 countries are producing cotton, which provides raw materials to the industry and employment opportunities for the people. Limiting the cost of production, conserving the ecosystem, and improving the cotton yield are key ingredients of sustainable cotton production. The inception of transgenic cotton (Bt varieties) improved yields, curtailed pesticide uses and promoted environmental safety. Over the years, Bt varieties of cotton have lost resistance against bollworms, and yield is declining. Unwise application of pesticides is associated with the development of resistance to sucking pests of cotton. The present review was conducted to undertake (i) the background of cotton production in the world, (ii) varietal development in cotton, (iii) problems (increasing susceptibility) in Bt varieties (iv) reliance on pesticides, and (v) myths of pesticides applications followed by pesticides knowledge among farmers. The cotton varieties which are resistant against sucking pests should be introduced. In addition, the promotion of biopesticides and fostering the adoption of Integrated Pest Management Approaches could be effective for the management of sucking pests. Farmers must be trained in site-specific and accurate spraying techniques by agricultural extension and plant protection departments. Accurate spraying techniques will not only improve pest control but also help in curtailing the environmental pollution being caused by the excessive use of pesticides in cotton.

Keywords: Sucking Pests, Resistance, Susceptibility, Bollworms, Ecosystem, Biopesticides.

1. INTRODUCTION

The world is producing around 25 M tons of cotton. India, China, the United States, Pakistan, Brazil, Australia, Uzbekistan, Turkey, Turkmenistan and Burkina Faso are the top ten cotton-producing nations in the world [1]. Globally, the total cotton production was expected to reach 121.6 million bales in 2021-22 [2]. *Gossypium hirsutum*, *Gossypium babadense*, *Gossypium arboretum*, and *Gossypium herbaceum* are four commonly grown types of cotton. A major proportion of the cotton area is devoted to Bt cotton [3]. The global cotton production is forecasted to grow by 1.6% and reach 30.6 million tons by 2031. The average yield of cotton is also to grow at 1.3%, which is likely to back the increase in global production of cotton [4].

Cotton production is much needed for the world for many reasons, including an addition to the economy and support to livelihoods [5].

Conventional cotton was not very effective due to severe susceptibility to pests. Transgenic cotton helped to combat the pest's attack. Since the inception of transgenic cotton, pesticide use patterns have changed, production increased, and the natural enemy population jumped high, followed by improved land use efficiency and productivity [6]. However, transgenic cotton has started showing susceptibility towards pests; moreover, pests have developed resistance. This transition is not only decreasing production but also increasing the cost of pesticide application along with the occurrence of environmental degradation [7]. The improper application of pesticides seems more critical in developing resistance and leading transgenic cotton to failure.

With a population of over 212.82 million and a population growth rate of 2.4%, Pakistan is the 6th most populous nation. Agriculture contributes to 18.5% of the country's GDP and provides

livelihood support to 38.5% of the population, making it a crucial sector for the country [8]. Cotton is one of the most significant crops produced in Pakistan. Cotton contributes to national economic development and for supporting the livelihoods of millions of farm families. Cotton has tremendous export potential, accounting for 55% of all foreign exchange incomes in Pakistan. Of the total farmers, 26% grow cotton in Pakistan, whereas 15% of the total cultivated area is devoted to cotton cultivation [9]. Approximately 65% of cotton is grown in Punjab province, which has arid conditions that are suitable for the crop. The remaining cotton is cultivated in Sindh, where the climatic conditions are relatively more humid [9]. Negligible areas in Khyber Pakhtunkhwa and Baluchistan are devoted to cotton. Cotton production contributes 4.5% of agriculture's value and 0.8% of the total GDP [10]. The area and production of selected countries using Bt varieties of cotton are presented in Figures 1 and 2. Figure 1 shows that the India, USA, Pakistan, China, Brazil and Australia are the leading countries in terms of cotton area devoted to Bt varieties of cotton [11]. Figure 2 shows that China, India, Pakistan, Brazil, Turkey and Uzbekistan were the top cotton-producing countries [12].

It is necessary to support cotton production, reduce the cost of production, improve quality, and implement policies to make farming profitable. Despite this, the agriculture sector has remained underdeveloped, and enhancing its performance is crucial for economic growth and poverty reduction. The present review is an integrated review to explore the cotton landscape in the world under prevailing conditions of Insect Pests and diseases. Effective management of insect pests is essential as they play a significant role in reducing the cotton

yield. Over time, the insects' pests have gained resistance; this resistance is associated with the unwise application of chemicals. This review aimed to endorse accurate spraying techniques to manage the infestation of insect pests and conserve the ecosystem. This review was initiated from the (i) background of cotton production in the world, (ii) varietal development in cotton, (iii) problems (increasing susceptibility) in Bt varieties (iv) reliance on pesticides and (v) myths of pesticides applications followed by pesticides knowledge among farmers.

2. VARIETAL DEVELOPMENTS IN COTTON

2.1. Inception and Promotion of Bt Cotton

Conventional cotton was predominantly the critical choice of farmers before the inception of transgenic cotton. Over the period, traditional cotton was perceived as less effective for different reasons. Of the major reasons, low production was the foremost. Infestation of pests, especially armyworms, was intensive on conventional cotton, resulting in the high cost of production and exhibiting less production. Abro *et al.* [13] identified that cotton bollworms, mainly American, Pink, and Spotted bollworms, were the offensive pests, bringing around 30-40% yield loss. Farmers had to apply extensive chemicals to control the bollworms, which not only skyrocketed the cost of production but also accelerated the resistance of bollworms. PACRA [14] reported that during the fiscal year 2022, Pakistan imported pesticides worth USD ~201.7 mln, and ~69% of it was consumed on cotton crop only. In this context, conventional

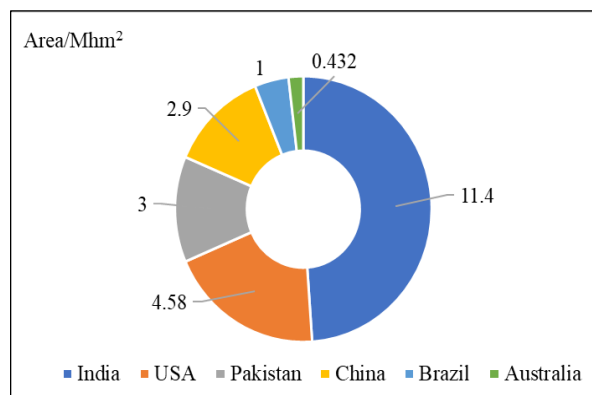


Fig. 1. Bt cotton cultivated area (Mh) in top cotton growing nations [11].

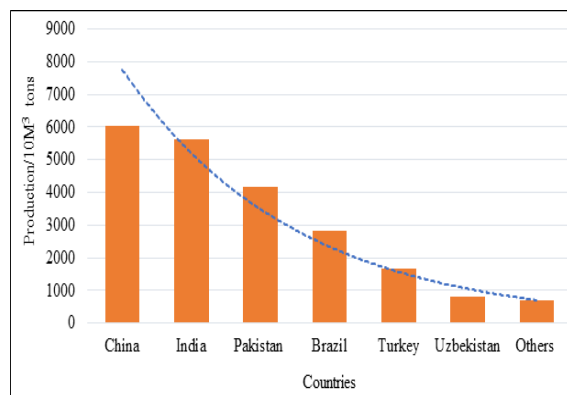


Fig. 2. Country-wise production of cotton in 2018-19 (1000 metric tons) [12].

cotton was deemed important for upgrading to new germplasms that are likely resistant to bollworms. The Pakistani government, in collaboration with the Center of Excellence in Molecular Biology (CEMB) and the National Institute for Biotechnology and Genetic Engineering (NIBGE), invested significant resources and human resources to develop genetically modified local varieties, specifically to tackle the issues in cotton production [15].

The first Bt variety of cotton was informally grown in Pakistan in 2002 [16]. However, the Bt variety obtained the final approval of the Government of Pakistan for cultivation in 2010. Contrary to Pakistan, the USA approved and commercialized the first Bt cotton variety in 1996 [17]. James [18] found that 12.1 million farmers had adopted GMO (Genetically Modified Organisms) cotton, with the majority of them in China and India. In Pakistan, some private seed companies introduced the first Bt variety seed in the district Rahim Yar Khan [16].

The average yield of cotton in Pakistan was 0.5 t ha⁻¹ considerably lower than the average yield achieved in China 9 t ha⁻¹, indicating a wide yield gap in Pakistan [19]. The inception of the Bt variety was deemed important in bridging this yield gap of cotton prevailing in Pakistan [19]. The details of Bt and Non-Bt varieties approved in Pakistan from 2010-19 are tabulated in Table 1 [20].

2.2. Performance of Bt Varieties

Bt varieties were aimed at curtailing the undue application of pesticides and allowing growers to control insects and pests through minimum use of pesticides. Some benefits of Bt varieties are less attack of bollworms, minimum use of pesticides, high yield potential, and reduction in labour, which lure farmers to adopt Bt varieties over the non-Bt varieties [21]. Studies such as Qaim *et al.* [22], Bennett *et al.* [23], Subramanian and Qaim [24] and Kouser and Qaim [25] have confirmed that

Table 1. Bt and Non-Bt varieties approved in Pakistan from 2010-2019 (Adapted from Razzaq *et al.* [20]).

Sr. no.	Variety name	Year of release	Sr. no.	Variety name	Year of release
Bt varieties			30	IUB-13	2015
1	IR-3701	2010	31	FH-LALAZAR	2015
2	Neelum-121	2010	32	BS-52	2015
3	FH-113	2010	33	BH-184	2015
4	Sitara-008	2010	34	Cyto-177	2015
5	MG-6	2010	35	AGC-999	2015
6	Ali Akbar-703	2010	36	MNH-988	2015
7	Ali Kabar-802	2010	Non-Bt varieties		
8	IR-1524	2010	37	Cyto-124	2015
9	GN Hybrid-2085	2010	38	NIAB-2008	2015
Non-Bt varieties			39	GOMAL-105	2015
10	Sindh-1	2010	Bt varieties		
11	CRIS-342	2010	40	CIM-602	2016
12	NIA-Ufaq-08	2010	Non-Bt varieties		
13	Malmal	2010	41	NIAB-KIRAN	2016
Bt varieties			42	CIM-620	2016
14	Tarzan-1	2012	43	CRIS-129	2016
15	MNH-886	2012	44	RJ-120	2016
16	NS-141	2012	Bt varieties		
17	FH-114	2012	45	IR-NIBGE-4	2017
18	IR-NIBGE-3	2012	46	IR-NIBGE-6	2017
19	CIM-598	2012	Non-Bt varieties		
20	Sitara 009	2012	47	RH-668	2018
21	A-One	2012	48	CIM-632	2018
Non-Bt varieties			49	NIAB-1048	2018
22	CIM-573	2012	50	NIAB-545	2018
23	FH-941	2012	51	Crystal-12	2018
24	FH-942	2012	52	Sitara-15	2018
25	GS-1	2013	53	Sahara-150	2018
Bt varieties			54	CIM-610	2018
26	AGC-777	2015	55	RH-662	2018
27	MM-58	2015	56	FH-152	2018
28	LEADER-1	2015	57	FH-490	2019
29	VH-305	2015			

with increased adoption of Bt cotton pesticide use decreased and cotton yield increased. As a result of the adoption of Bt varieties, not only was the use of pesticides reduced, but the cost of production also significantly declined, and the incidences of poisoning were reported to decrease [25]. USA [26], Mexico [27], South Africa [28], China [29], India [30] and Pakistan [21] have witnessed the benefits of Bt cotton, and its adoption was positive for the farmers for economic reasons as the yield was higher than the conventional cotton varieties [31]. Specifically, small farmers' yield increased due to Bt cotton adoption [32, 33]. Hofs *et al.* [34] reported that the economic returns of the Bt adoption were very bright, primarily due to a significant reduction in pyrethroid production. The overview of income benefits obtained from adopting Bt varieties is portrayed in Figure 3. This shows that India, China, USA, Pakistan, Brazil, Australia and Argentina were leading countries harvesting noteworthy income through genetically modified cotton [35].

3. PROBLEMS (INCREASING SUSCEPTIBILITY) IN BT VARIETIES

Bt varieties had resistance against the bollworms [13]. Despite its effectiveness, Bt cotton did not show high efficacy against insects that suck sap, such as whitefly, jassid, thrips, and mealybugs [36]. Dutt [37] reported a high infestation of mealybugs in Bt cotton varieties. Over time, Bt varieties are becoming vulnerable to pests and diseases, and their resistance is decreasing. Bt cotton contains a gene from the Bt bacterium that produces endotoxins harmful to certain Lepidopteran pests, including the Pink Bollworm [38]. However, the unforeseen

return of key cotton pests that Bt was aimed to resist has augmented a new debate to rethink the resistance level of Bt cotton [39, 40]. Different pests attack Bt in various varieties, like different bollworms and whiteflies. In the subsequent section, debate is made about how bollworms, sucking pests, insects and viruses are influencing Bt varieties.

3.1. Bollworms

Bt varieties have resistance against the bollworms, which were more hazardous in lowering the production and quality of cotton. Karar *et al.* [41] admitted that the risk of bollworms was significantly reduced in cotton after the introduction of Bt varieties. However, over the period, the resistance in Bt varieties decreased. Rabelo *et al.* [42], Zhang *et al.* [43] and Tabashnik *et al.* [44] reported that the Bt resistance against the bollworms was decreasing. The attack of Pink Bollworms on Bt cotton is more often visible [41]. Pink Bollworm (PBW) infestation on Bt varieties surprised the growers, and this attack caused a severe loss in yield and income [45]. The results of Rajput *et al.* [46] were confirmatory that PBW caused a reduction in the yield and caused damage to cotton lint quality, producing yellow spots on the fibre. This infected lint got lower rates in the market and significantly lowered the farmers' income.

PBW was also found to be responsible for delaying the maturity of cotton [47]. Studies were evident that PBW attacked all varieties of cotton. For instance, Gutierrez *et al.* [48] identified that PBW damaged the cotton crop grown in rain-fed condition and damage was relatively more serious as compared to the cotton grown in irrigated conditions. This damage was associated with climatic variations. However, they questioned the use and quality of Bt cotton seed and the number and quality of insecticides applied, especially in rainfed cotton. The findings of Lu *et al.* [49] were different to the studies conducted in Pakistan and India, as they found that Bt cotton varieties were effectively capable of controlling the population of bollworms in China. Resistance management methods are employed to control bollworms, significantly to alleviate the host plants and improve the resistance in Bt varieties. Rajapakse and Walter [50] found more attacks of bollworms in the cotton field as many weeds were growing around the field. In addition to field cleaning practices, the cultivation

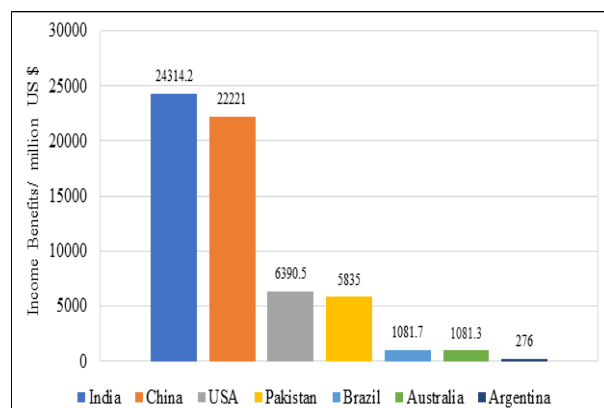


Fig. 3. Income benefits of genetically modified (GM) cotton farm in selected countries, 1996–2016 (million US\$) [35].

of natural refuge crops has been identified as a successful strategy to postpone the development of resistance to cotton bollworm [51]. It was recommended to grow non-Bt varieties alongside Bt varieties as a means of sufficiently delaying the development of bollworm resistance to Bt [52].

3.2. Cotton Leaf Curl Virus (CLCv)

The Gemini virus is responsible for causing Cotton Leaf Curl Disease (CLCv), which is a severe issue in cotton-producing regions worldwide. This virus spreads through the whitefly (*Bemisia tabaci*), and it can devastate the crop, leading to up to a 90% reduction in yield. This can also deteriorate the fibre quality [53]. The initial outbreak of CLCv was observed in Nigeria in 1912, and it has since spread to various cotton-producing countries, including the United States of America, Pakistan, India, and China [54]. Sattar *et al.* [55] argued that India and Pakistan were facing whitefly infestation in the cotton crop, resulting in losing the resistance. In 1989, CLCv was detected in the experimental fields of the Indian Agricultural Research Institute, New Delhi. Similarly, the existence of CLCv was later seen in Karnataka state, South India [56], and Northwest India in 1994 [57]. Specifically in

Pakistan, the CLCv was first reported in Multan in 1967, and was perceived as a minor disease till 1987. However, it has spread over one million hectares of cotton area over the years as presented in Table 2 [58, 59]. CLCv damaged approximately 60% of the cotton crop in Multan in 1988, and the damage continues to occur each year. Pakistan and India have seen two epidemics of CLCv attack on cotton, causing a significant loss in production [55].

The incidences of CLCv are constantly changing over time, pointing towards the likely increase in yield decline [60]. CLCv grows in a suitable environment. Whitefly vectors can expedite the mode of spread [55]. The epidemiology of CLCv disease was related to abiotic factors like temperature and age of the plant [61]. Amrao *et al.* [62] found CLCv a very prominent disease. Around 70% of yield decline was witnessed subject to CLCv. The adverse impacts of the CLCv were more prominent on ginning out-turn and fibre fitness [63]. Nazeer *et al.* [64] stated that none of the varieties of *G. hirsutum* had resistance against the CLCv, although *G. Arboreum* has resistance against CLCv. In another study, Nawaz *et al.* [61] suggested changing the sowing times, cultural techniques, crop nutrition, buffer crops, whitefly

Table 2. Important landmarks in the history of CLCv. (Adapted from Rahman *et al.* [54]).

Year	History of CLCv
1912	<ul style="list-style-type: none"> • First report of CLCUD in Nigeria
1924	<ul style="list-style-type: none"> • Spread in Africa
1967	<ul style="list-style-type: none"> • CLCuD reported in Pakistan
1990	<ul style="list-style-type: none"> • First epidemic in Pakistan • Characterization of CLCuD initiated
1997-99	<ul style="list-style-type: none"> • Discovery of DNA alpha-satellite in 1999 • CIM-1100 was the first resistant variety followed by a number of varieties including CIM 448, CIM-496, NIBGE-2, FH-901, FH-1000, MNH-552, etc released in subsequent years
2001	<ul style="list-style-type: none"> • In 2001, CLCuBuV appeared in Pakistan • Beta-satellite discovered • DNA marker associated with resistance were identified
2005-10	<ul style="list-style-type: none"> • Antisense RNA, RNAi, etc has been utilized • Introgression breeding started • Release of tolerant cotton genotypes (NIBGE-115 and NN-3) • CLCuD reported in China
2012-13	<ul style="list-style-type: none"> • Identification of asymptomatic cotton accession • Utilization in breeding as well as for DNA marker identification
2016	<ul style="list-style-type: none"> • Development of asymptomatic cotton lines • Identification of QTLs associated with diseases resistance

control and use of systemic chemicals followed by the seed treatment to manage the CLCv outbreak. Contrarily, late sowing of cotton, excessive use of nitrogenous fertilizer, and more attacks on insect pests exuberated the CLCv [65].

3.3. Sucking Insect Pests

Insects that suck sap, such as whitefly, thrips, and aphids, can be harmful to the health and productivity of cotton plants. Sucking insect pests have swapped the cotton bollworms. Since the inception of Bt varieties of cotton, about a 97% decrease in the chemical application was recorded. In the meantime, insecticides application jumped by 154% to control the sucking insect pests [66]. The overuse of insecticides had negative environmental consequences. Bt cotton varieties were not adequately resistant to insects that suck sap [67]. Therefore, continuously applying pesticides and insecticides is necessary to manage insect pests [34]. If pesticides are not appropriately applied, the population of insect pests multiplies, with significant inverse repercussions on the crop [68].

Several studies like Abro *et al.* [13], Naveen *et al.* [69] and Sun *et al.* [70] augmented the higher infestation of sucking pests, including thrips, jassid, and whitefly, causing a significant decline in the cotton yield [71]. Jassid, thrip and Whitefly sucked the sap from the plant, turning the plant weak and ending in wilting and shedding of the leaves. These sucking pests have specific damage at the seedling stage of the crop and vegetative growth stage [13]. Whitefly further exaggerated the loss causing cotton leaf curl virus (CLCv) on cotton crops. India, one of the leading cotton producers, has reported a severe attack of whitefly on cotton during 2015-16 [69]. Sucking pests had a direct association with temperature and rainfall. Murtaza *et al.* [72] depicted that climate change had a significant impact on the population of sucking pests like Jassid. The findings of Harde *et al.* [73] confirmed that there was a positive correlation between the occurrence of sap-sucking pests such as whitefly, Jassid, Thrips, Mealybug, and Aphid and the maximum temperature, minimum temperature, and maximum relative humidity. In another study, Shahid *et al.* [74] pointed out that the thrips and mites' population was high in June, August and September, augmenting the Jassid, whitefly and mealy bug population. This accentuates that with

climate change, there are more chances of a high population of sucking pests. Therefore, with the increase in population, the chances of cotton yield losses will remain higher.

4. RESISTANCE DEVELOPMENT IN PESTS

Pertinent to increased resistance, the efficacy of Bt cotton varieties has declined [75]. Different factors contribute to the development of resistance to insect pests. Noncompliance with environmental protection agency regulations, inadequate production of high Bt endotoxins, repeated exposure to the same Bt endotoxins, and cross-resistance to multiple Bt endotoxins [76]. The variation in resistance of cotton armyworms and bollworms to insecticides was linked to the varying expression levels of Cry1Ac in the field conditions. This expression level was influenced by the plant's variety, age, and environmental factors [77, 78]. The resistance of insect pests was also found to increase due to insecticidal efficacy. The effectiveness of Bt varieties as insecticides was uneven due to the fluctuating expression of Bt protein throughout the cotton-growing season [79]. Insecticidal efficacy and ability were directly or indirectly predisposed by insect pests, disease intensity, rainfall amount, soil attributes, and apposite farm management. It can be deduced that an optimal environment was obligatory for Genetically Modified (GM) Cotton, which eventually reinforced the Bt gene expression. Huang *et al.* [76] specifically cited that in India, the development of resistance in pink bollworm (PBW) took seven years, which was attributed to the cultivation of unauthorized Bt cotton seeds. This resistance was associated with the low dose of Bt protein and non-compliance with the refuge strategy. Luo *et al.* [80] and Sethi and Dilawari [81] agreed that resistance in cotton insects and pests was due to the unwise application of pesticides on the crop. Sparks and Nauen [82] reported serious resistance in *B. tabaci* (whitefly), which is known for its enormous damage to the cotton crop in particular. The increased resistance in whiteflies has been reported in some countries [83, 84], and intensive pesticide use for insect and pest management inflated the infestation of whiteflies [85]. Ahmad *et al.* [63] reported a moderate resistance of whitefly against pyrethroids. It can be deduced that resistance to insect pests is increasing, and the unwise application of pesticides remains the critical reason.

4.1. Reliance on Pesticides Application

According to the report of Khan *et al.* [86], A total of 4875 kg of pesticide active ingredients were applied by farmers in the cotton belt of Punjab, Pakistan, per annum. The import of pesticides is also increasing each year [87]. Around 80% of all pesticides are used on cotton crops, while the remaining 20% are used on other crops like rice, sugarcane, maize, wheat, fruit orchards, and vegetables [87]. In the last 20 years, an expansion of 11.69% in the application of pesticides has been witnessed in Pakistan, and the total number of sprays per crop has reached more than 10 [88]. This extensive application is a point to ponder as human health is vulnerable to the impacts of chemicals, and insect pests have developed a high resistance level against the chemicals. No wonder the application of pesticides showed an increase in production by controlling the infestation of insect pests. From 1995 to 1998, an unexpected outbreak of insect pests caused a significant reduction in cotton yield from 849 to 230 kg/ha [89]. To combat this problem, the Government of Pakistan and pesticide manufacturers implemented a program involving increased pesticide use for monoculture crops. As a result, cotton production substantially increased from 2000 to 2001 [90]. The average yield in 1999-2000 reached 643 Kg/ha, from 494 Kg/ha, as recorded in 1997-98 and 1998-99. However, farmers in cotton-growing regions were applying 8 to 13 sprays per season and exceeding the recommended dosage to combat insect pests of cotton [91].

The extensive utilization of insecticides for managing whiteflies and other co-existing pests has resulted in a significant reduction of its natural enemies and the development of resistance to most of the conventional insecticides, thereby largely triggering whitefly outbreaks [63, 92]. According to the Central Cotton Research Institute (CCRI) in Multan, the two main cotton pests, the American bollworm and the whitefly, have developed resistance to commonly used pesticides [63, 92]. According to the Ministry of Food and Agriculture's report, there is evidence of a pesticide treadmill in Pakistan, as seen by the significant increase in applications of monocrotophos, cypermethrin, methamidophos, and dimethoate for controlling pests such as the American bollworm and whitefly. The application of monocrotophos has increased by 19 to 720 times, cypermethrin by 26 to 168-fold,

methamidophos by 40 to 492-fold, and dimethoate by 104 to 725-fold to control these pests. Tariq *et al.* [91] indicated that before 1983, only a small percentage (5 to 10%) of cotton-growing regions in Punjab were treated with pesticides. However, this percentage increased to 100% by 1997. Overusing pesticides leads to insect pests developing resistance and poses a risk to farmers and community health. Excessive use of pesticides, such as spraying more than the recommended dose, in cotton fields poses a significant risk to field workers and pickers. It may result in unacceptable residue concentrations in cottonseed oil and cakes [90]. The discussion on the use of pesticides suggests that while their application has both positive and negative effects, the outcome depends largely on the spraying methods used by farmers. Further discussion on the various spraying techniques follows.

4.2. Myths of Spraying Techniques

Spraying techniques are of great worth in the wake of getting effective control over insect pests. In this regard, farmers use different spraying techniques. According to Tahir *et al.* [93], farmers typically use Knapsack and tractor-mounted boom sprayers to apply pesticides to crops in Pakistan. However, a significant issue with these sprayers is that the nozzle pressure is not consistently maintained during application, resulting in pesticide loss due to dribbling or drift. These problems increase production costs and contribute to environmental pollution and ecosystem imbalances. Damalas and Eleftherohorinos [94], most of the pesticides applied to the crops are wasted when they are used by farmers using improper techniques and nozzles. This wastage is an extra addition to the cost of production indeed.

Proper use of spraying techniques and nozzles greatly contributed to combatting insect pests. The type of nozzle, size, pressure on droplet size, and velocity are some aspects that farmers should consider palatably while spraying. The concerns made in this regard not only save the quantity of the pesticide but also curtail the cost of production [95]. Nozzles are classified based on droplet size and spray pattern [96]. Nozzles have three significant roles: breaking down the liquid into small droplets, dispersing these droplets in a specific pattern, and controlling the sprayer's release rate. Unfortunately, farmers have a poor

understanding of nozzle function and often use too much or too little pesticide, leading to ecological disruption and pesticide waste [97]. Due to untrained pesticide applicators and faulty spray equipment, many pesticides sprayed on crops reach non-target areas [97]. Excessive use of pesticides can result in the death of unintended organisms, leading to the potential resurgence of insect pests [98]. Insecticides have become less effective against certain cotton pests, such as *Heliothis* and whitefly, due to the inappropriate use of pesticides [99, 100]. Ejaz *et al.* [101] augmented that approximately 50% of the pesticides applied to various crops are wasted on non-targeted areas, leading to increased production costs and environmental degradation of the surrounding soil, water, and air. This is mainly due to ineffective spraying machines that cannot maintain the specified nozzle pressure, discharge, and height, which impacts spray pattern, droplet size, and uniformity. The size of droplets can also affect the spray coverage, and as droplet size decreases, the potential for drift increases [102]. Klein *et al.* [103] pointed out that boom sprayers' non-uniform nozzle tip output resulted in a misapplication of pesticides. The components of a nozzle include a nozzle body, a strainer, a replaceable nozzle tip, and a cap for securing it. Although farmers in Iran have excessively used chemical pesticides, there is little comprehension of their criteria for selecting and utilizing pesticides [104]. Ensure the use of pesticides correctly by paying attention to the pressure and the spraying situation before application [105]. Improper use of spraying techniques harnesses its damage to human health as well. While developing countries use only 20% of the world's total pesticide volume, they suffer from disproportionately high pesticide poisoning rates [106]. This pity situation spotlights a dire need to combat the issue of improper handling and use of pesticides in the wake of effective ecosystem conservation and escalated knowledge levels to combat the issue of improper use of pesticides. In this regard, knowledge regarding pesticide application is regarded as mandatory.

4.3. Dilemmas of Pesticide Knowledge

Among various factors behind the ineffective use of spraying techniques and wastage of resources, inadequate knowledge on the part of farmers is more prominent. Knowledge and information about the appropriate use of pesticides are regarded

as key to the effective use of pesticides [107, 108]. Farmers in Pakistan usually lack the knowledge to select appropriate pesticides and effective spraying techniques as per their needs. Allahyari *et al.* [109] reported that farmers moderately understood precise spraying techniques. Most of the farmers preferred pesticides that were easy to access from dealers and used in the field [110]. Ejaz *et al.* [101] reported a lack of knowledge regarding the detrimental consequences of pesticide exposure during pesticide application. Various researchers such as Lekei [111], Ngowi [112], Karamidehkordi and Hashemi [113] have reported that farmers' limited education levels and inadequate training of pesticide management have resulted in their insufficient knowledge about the appropriate use of pesticides in suitable amounts. Farmers had limited adoption of recommendations for the safe use of pesticides due to inadequate technical awareness and knowledge [114]. Farmers rely on traditional sources to acquire information rather than contacting technical institutions like plant protection and agriculture extension departments. Khan and Iqbal [115] reported farmers' over-reliance on fellow farmers for information as 70% of farmers obtained guidance on the safe use of pesticides from their fellow farmers. Rehman *et al.* [116] reported that fellow farmers were reported to be a primary source of guidance for selecting and using pesticides, including information on their proper handling and usage. In another study, Mubushar *et al.* [114] revealed that farmers were not firmly following the advice and recommendations disseminated by the extension field staff regarding the safe use of pesticides. The over-reliance of farmers on fellow farmers to acquire information has already been reported by various research studies such as Ashraf *et al.* [117] and Akinagbe *et al.* [118].

5. IMPLICATIONS FOR THE WISE APPLICATION OF PESTICIDES AND INSECT PEST MANAGEMENT

Achieving higher production is the foremost concern for farmers and state institutions. For this reason, farmers implement whatever possible measure is needed. Among various options, the application of pesticides, insecticides and fungicides is perceived prominent as an infestation of insects, pests and diseases on crops not only significantly lower the level of production but also deteriorates the quality of produce. Tariq *et al.* [91] reported that a hefty pest

attack during 1995-1998 reduced cotton production from 849 kg/ha to 230 kg/ha. Mealybug infestations in Pakistan was expected to decrease cotton yield by 1.3 million bales [119]. Mallah *et al.* [120] and Abbas *et al.* [121] reported a 20-40% loss in cotton production due to different insects and pests of the cotton. Around 24% of cotton production loss was due to jassid [20].

The implications for applying pesticides and insect pest management in cotton crop cultivation are paramount for ensuring sustainable and productive agriculture. Cotton, a major cash crop, is susceptible to various insect pests that can significantly impact yields if uncontrolled. Wise application of pesticides involves a judicious use of chemical agents, considering factors such as pest species, developmental stages, and potential ecological consequences. Wise application of pesticides in the cotton crop can reduce weed-related yield reduction by 50 to 85% and increase production costs by addressing labour shortages and labour wages [122]. Although farmers use various pest control methods to increase production and reduce yield loss, no single method of controlling insect pests is considered sufficient to achieve the desired production level. Using chemicals to protect crops is beneficial and an essential component of integrated pest management [123]. The integration of control measures can increase production significantly. If cotton pests are effectively controlled, the yield can be increased by 200-300 kg per hectare [124]. The government of Pakistan and pesticide manufacturers launched a program aimed at increasing the use of pesticides for monoculture crops. As a result, cotton production experienced a significant boost from 2000 to 2001 [89].

Integrated Pest Management (IPM) strategies, encompassing biological, cultural, and chemical control methods, emerge as a holistic approach to curb pest populations while minimizing environmental impact. Adopting IPM addresses immediate pest concerns and contributes to long-term pest resistance management, preserving the efficacy of available pesticides. Integrated pest management (IPM) in cotton crops can lead to long-lasting, economical, and eco-friendly benefits [125]. In a study, Kranthi and Russell [126] revealed that Integrated Pest Management (IPM) programs, using naturally occurring pest control components, result in favourable ecological, sociological, and

environmental consequences for the cotton crop. The implications for the wise application of pesticides extend beyond pest control to broader environmental and economic dimensions. Overreliance on certain pesticides can lead to the development of resistant pest populations, necessitating a rotation of active ingredients to maintain efficacy. Moreover, indiscriminate pesticide use can adversely affect non-target organisms, soil health, and water quality [94]. A wise and informed approach involves selecting pesticides with lower environmental impact, adhering to recommended application rates, and incorporating non-chemical pest management methods whenever possible [127, 128]. Economically, the judicious use of pesticides minimizes production costs and conserves resources. By investing in research and extension services that promote wise pesticide application and integrated pest management, the agricultural sector can strike a balance between effective pest control, environmental stewardship, and economic sustainability in cotton cultivation [129]. Matteson [130] stated that Integrated pest management (IPM) extension education encourages wise pesticide use and empowers farmers to manage a healthy paddy ecosystem, promoting ecological balance in rice paddies. Improved agricultural extension services, such as the PlantWise program in China, lead to more sustainable pest management by increasing recommendations for biological control, pest monitoring, and cultural control [131]. Integrated pest management training in school extension programs effectively achieves sustainable agriculture [132]. Similarly, the Insecticide Resistance Management (IRM) program in India significantly improved farmers' knowledge in identifying insect pests, proper use of insecticides, and timely sowing of the cotton crop, but limited their understanding of cultural practices.

6. CONCLUSIONS AND RECOMMENDATIONS

Insect pests and diseases are key factors lowering the cotton yield. Insect pests in cotton crops are managed by applying various pesticides, which not only increase the cost of production but also contribute to environmental degradation. Over the years, insect pests have attained resistance against pesticides; even the resistance of the Bt gene has decreased being susceptible to bollworms (i.e. Pink Bollworm) and other sucking pests (i.e.,

whitefly). The unwise application of pesticides on cotton crops is the foremost reason for increasing resistance to insect pests. Cotton growers are usually unaware of the technical side of the spraying. Using traditional spraying methods was the ineffective and injudicious application of pesticides that caused environmental pollution. Farmers are deficient in technical knowledge about the safe application of pesticides. The deficiency of technical knowledge is also associated with reliance on traditional information sources (i.e., fellow farmers). Therefore, it is essential to improve farmers' technical knowledge of spraying techniques and curtail the cost of production. Helping farmers in pesticide selection, controlling overdosing/underdosing, and conserving the ecosystem, environment, soil and water should be priorities. Advisory service-providing institutions should technically guide the farmers regarding the safe use of pesticides. Developing trained and technically sound farmers who can serve as an information source for other farmers seems another vital avenue to consider. Moreover, there is a need for the development of sucking pest-resistant cotton varieties followed by the implementation of integrated pest management techniques.

6.1. Direction for Future Research

Based on this critical review, the following future research directions are proposed;

1. Future research could explore and evaluate alternative pest management strategies that reduce reliance on traditional spraying techniques. This might involve the assessment of biological control methods, such as the introduction of natural predators or the use of microbial agents to control sucking pests.
2. Research could focus on integrating precision farming technologies to optimize pesticide application in cotton fields. Precision agriculture tools, such as drone-based monitoring, satellite imaging, and sensor technologies, can potentially target pest-infested areas more precisely.
3. Understanding the factors influencing farmers' decision-making regarding pesticide use is crucial. Future research could delve into behavioural studies to explore the motivations, knowledge gaps, and perceptions contributing to unwise spraying techniques. Developing targeted educational interventions and extension

programs to enhance farmers' understanding of pest management practices, including the consequences of unwise spraying, could be an effective strategy to address the issue.

4. To comprehensively address resistance development in sucking pests, a longitudinal study could be conducted to monitor resistance patterns over an extended period. This research would involve regular sampling and analysis of pest populations in cotton fields to track changes in resistance levels. Understanding the dynamics of resistance development over time could inform the development of more sustainable pest management strategies.

7. CONFLICT OF INTEREST

The authors declared no conflict of interest.

8. REFERENCES

1. M.A. Khan, A. Wahid, M. Ahmad, M.T. Tahir, M. Ahmed, S. Ahmad, and M. Hasanuzzaman. World cotton production and consumption: An overview. In: Cotton production and uses. S. Ahmad, and M. Hasanuzzaman (Eds.). *Springer, Singapore* pp. 1-7 (2020).
2. L. Meyer, and T. Dew. Cotton and wool outlook: December 2022. *US Department of Agriculture, Economic Research Service* (2022). <https://www.ers.usda.gov/webdocs/outlooks/105429/cws-22k.pdf?v=5324.1>.
3. F. Shuli, A.H. Jarwar, X. Wang, L. Wang, and Q. Ma. Overview of the cotton in Pakistan and its future prospects. *Pakistan Journal of Agricultural Research* 31: 396-407 (2018).
4. OECD. OECD-FAO Agricultural Outlook 2022-2031. *Food and Agriculture Organization (FAO) of the United Nations* (2022). <https://www.oecd-ilibrary.org/docserver/f1b0b29c-en.pdf?expires=1722581785&id=id&accname=guest&checksum=5A136EEC5F26156B8A781223D0691956>.
5. M.Y. Ali, S. Saleem, M.N. Irshad, A. Mehmood, M. Nisar, and I. Ali. Comparative study of different irrigation system for cotton crop in district Rahim Yar Khan, Punjab, Pakistan. *International Journal of Agricultural Extension* 8: 131-138 (2020).
6. G. Brookes. Genetically Modified (GM) Crop Use 1996–2020: Environmental Impacts Associated with Pesticide Use Change. *GM Crops and Food* 13: 262-289 (2022).
7. S. Ahmad, S. Akhtar, S. Bhatti, S. Imran, M.S. Akhtar, G. Mustafa, A.R. Aslam, C. Liu, S. Noreen, and M.A. Khan. Assessment of the impact of climate change on the productivity of cotton: empirical evidence from cotton zone,

- southern Punjab, Pakistan. *International Journal of Agricultural Extension* 9:143-162 (2021).
8. Government of Pakistan. Economic Survey of Pakistan, *Finance Division Islamabad, Pakistan* (2018). https://www.finance.gov.pk/survey/chapters_19/Economic_Survey_2018_19.pdf.
 9. A.W. Rana, A. Ejaz, and S.H. Shikoh. Cotton Crop: A Situational Analysis of Pakistan. PACE Working Paper. *Pakistan Agricultural Capacity Enhancement Program (PACE), International Food Policy Research Institute – Pakistan (IFPRI)* (2020). <https://ebrary.ifpri.org/utils/getfile/collection/p15738coll2/id/133702/filename/133913.pdf>.
 10. Government of Pakistan. Economic Survey of Pakistan, *Finance Division Islamabad, Pakistan* (2009). https://www.finance.gov.pk/survey/chapter_10/02_Agriculture.pdf.
 11. ISAAA. Global status of commercialized biotech/GM crops: Brief No. 54. USA: *ISAAA NY*; (2018). <https://www.isaaa.org/resources/publications/briefs/54/>
 12. USDA-FAS. Cotton outlook 2019. <https://www.usda.gov/sites/default/files/documents/2019-aof-cotton-outlook.pdf>.
 13. G.H. Abro, T.S. Syed, G.M. Tunio, and M.A. Khuhro. Performance of Transgenic Bt Cotton Against Insect Pest Infestation. *Biotechnology (Faisalabad)* 3: 75-81 (2003).
 14. PACRA. Pesticides. *Sector study by Pakistan Credit Rating Agency* (2023). https://www.pacra.com/index.php/view/storage/app/Pesticides%20-%20PACRA%20Research%20-%20Feb%272023_1677423830.pdf.
 15. U. Shahbaz, X. Yu, and M.A. Naeem. Role of Pakistan government institutions in adoption of Bt cotton and benefits associated with adoption. *Asian Journal of Agricultural Extension, Economics and Sociology* 29: 1-11 (2019).
 16. M. Luqman, G.M. Shah, M.A.S. Raza, N. Shahid, and M. Hassan. Performance of Bt cotton varieties under Khanewal conditions. *Bulgarian Journal of Agricultural Science* 21: 105-108 (2015).
 17. C. James. Global Review of Commercialized Transgenic Crops: 2001 Feature: Bt Cotton. *The International Service for the Acquisition of Agri-biotech Applications* (2002). <https://www.isaaa.org/resources/publications/briefs/26/download/isaaa-brief-26-2002.pdf>.
 18. C. James. Global Status of Commercialized Biotech/GM Crops: *The International Service for the Acquisition of Agri-biotech Applications* Brief No. 39 (2008). <https://www.isaaa.org/resources/publications/briefs/39/executivesummary/pdf/Brief%2039%20-%20Executive%20Summary%20-%20English.pdf>.
 19. R. Ahsan, and Z. Altaf. Development, Adoption and performance of Bt cotton. *Pakistan Journal of Agricultural Research* 22: 73-85 (2009).
 20. A. Razzaq, M.M. Zafar, A. Ali, A. Hafeez, W. Batool, Y. Shi, W. Gong, and Y. Yuan. Cotton germplasm improvement and progress in Pakistan. *Journal of Cotton Research* 4: 1-14 (2021).
 21. M. Arshad, R.R. Khan, A. Aslam, and W. Akbar. Transgenic Bt cotton: Effects on target and non-target insect diversity. In: Past, Present and Future Trends in Cotton Breeding. M.U. Rahman, and Y. Zafar (Eds.). *IntechOpen* pp. 155-174 (2018).
 22. M. Qaim, A. Subramanian, G. Naik, and D. Zilberman. Adoption of Bt Cotton and Impact Variability: Insights from India. *Review of Agricultural Economics* 28: 48-58 (2006).
 23. R. Bennett, U. Kambhampati, S. Morse, and Y. Ismael. Farm-Level Economic Performance of Genetically Modified Cotton in Maharashtra, India. *Review of Agricultural Economics* 28(1): 59-71 (2006).
 24. A. Subramanian, and M. Qaim. The Impact of Bt Cotton on Poor Households in Rural India. *Journal of Development Studies* 46: 295-311 (2010).
 25. S. Kouser, and M. Qaim. Impact of Bt cotton on pesticide poisoning in smallholder agriculture: A panel data analysis. *Ecological Economics* 70: 2105-2113 (2011).
 26. F.J. Perlak, M. Oppenhuizen, K. Gustafson, R. Voth, S. Sivasupramaniam, D. Heering, B. Carey, R.A. Ihrig, and J.K. Roberts. Development and commercial use of Bollgard cotton in the USA - early promises versus today's reality. *The Plant Journal* 27: 489-501 (2001).
 27. G. Traxler, S. Godoy-Avila, J. Falck-Zepeda, and J. de Jesús Espinoza-Arellano. Transgenic Cotton in Mexico: A Case Study of the Comarca Lagunera. In: The Economic and Environmental Impacts of Agbiotech. N. Kalaitzandonakes (Ed.). *Springer Boston, MA, US* pp: 183-202 (2003).
 28. Y. Ismael, R. Bennett, and S. Morse. Farm-Level Economic Impact of Biotechnology: Smallholder Bt Cotton Farmers in South Africa. *Outlook on Agriculture* 31: 107-111 (2002).
 29. C.E. Pray, J. Huang, R. Hu, and S. Rozelle. Five years of Bt cotton in China - the benefits continue. *The Plant Journal* 31: 423-430 (2002).
 30. M. Qaim, and D. Zilberman. Yield Effects of Genetically Modified Crops in Developing Countries. *Science* 299: 900-902 (2003).
 31. V.P. Gandhi, and N. Namboodiri. The adoption and economics of Bt cotton in India: Preliminary results from a study. *Indian Institute of Management Ahmedabad, India* W.P. No. 2006-09-04 (2006). https://www.iima.ac.in/sites/default/files/rnpfiles/2006-09-04_vgandhi.pdf.
 32. A. Ali, and A. Abdulai. The Adoption of Genetically

- Modified Cotton and Poverty Reduction in Pakistan. *Journal of Agricultural Economics* 61: 175-192 (2010).
33. R. Mulwa, D. Wafula, M. Karembu, and M. Waithaka. Estimating the potential economic benefits of adopting Bt cotton in selected COMESA countries. *AgBioForum* 16: 14-26 (2013).
 34. J.L. Hofs, M. Fok, and M. Vaissayre. Impact of Bt cotton adoption on pesticide use by smallholders: A 2-year survey in Makhatini Flats (South Africa). *Crop Protection* 25: 984-988 (2006).
 35. G. Brookes, and P. Barfoot. Environmental impacts of genetically modified (GM) crop use 1996–2018: impacts on pesticide use and carbon emissions. *Crops and Food* 11: 4215-41 (2020).
 36. H.C. Sharma, and G. Pampapathy. Influence of transgenic cotton on the relative abundance and damage by target and non-target insect pests under different protection regimes in India. *Crop Protection* 25: 800-813 (2006).
 37. U. Dutt. Mealybug takes away glory of Bt cotton (2007). <http://www.ens-newsire.com/ens/aug2007/2007-08-24-insdutt.asp>.
 38. S. Kaviraju, D. Kumar, N. Singh, and S. Kumar. A Comparative Study on Socio Economic Impact of Bt cotton and Non-Bt cotton Farm Households in Warangal District of Telangana State. *International Journal of Current Microbiology and Applied Sciences* 7: 1561-1567 (2018).
 39. B.B. Fand, V.S. Nagrare, S.P. Gawande, D.T. Nagrale, B.V. Naikwadi, V. Deshmukh, N. Gokte-Narkhedkar and V.N. Waghmare. Widespread infestation of pink bollworm, *Pectinophora gossypiella* (Saunders) (Lepidoptera: Gelechiidae) on Bt cotton in Central India: a new threat and concerns for cotton production. *Phytoparasitica* 47: 313-325 (2019).
 40. K. Najork, S. Gadela, P. Nadiminti, S. Gosikonda, R. Reddy, E. Haribabu, and M. Keck. The Return of Pink Bollworm in India's Bt Cotton Fields: Livelihood Vulnerabilities of Farming Households in Karimnagar District. *Progress in Development Studies* 21: 68-85 (2021).
 41. H. Karar, M.J. Arif, M. Arshad, A. Ali, and Q. Abbas. Resistance/susceptibility of different mango cultivars against mango mealybug (*Drosicha mangiferae* G.). *Pakistan Journal of Agricultural Sciences* 52(2): 367-377 (2015).
 42. M.M. Rabelo, J.M.L. Matos, S.M. Orozco-Restrepo, S.V. Paula-Moraes, and E.J.G. Pereira. Like Parents, Like Offspring? Susceptibility to Bt Toxins, Development on Dual-Gene Bt Cotton, and Parental Effect of CryI_{Ac} on a Nontarget Lepidopteran Pest. *Journal of Economic Entomology* 113: 1234-1242 (2020).
 43. H. Zhang, W. Yin, J. Zhao, L. Jin, Y. Yang, S. Wu, B.E. Tabashnik, and Y. Wu. Early warning of cotton bollworm resistance associated with intensive planting of Bt cotton in China. *PLoS One* 6: e22874 (2011).
 44. B.E. Tabashnik, K. Wu, and Y. Wu. Early detection of field-evolved resistance to Bt cotton in China: Cotton bollworm and pink bollworm. *Journal of Invertebrate Pathology* 110: 301-306 (2012).
 45. K. Najork, J. Friedrich, and M. Keck. Bt cotton, pink bollworm, and the political economy of sociobiological obsolescence: insights from Telangana, India. *Agriculture and Human Values* 39: 1007-1026 (2022).
 46. I.A. Rajput, A. M. Lodhi, T.S. Syed, G.H. Abro, and I. Khatri. Comparative Biology of Pink Bollworm, *Pectinophora gossypiella* Saund. on Bt. and Non-Bt. Cotton. *Biological Sciences - PJSIR* 62: 116-121 (2019).
 47. J. Gore, B. Leonard, G. Church, and D. Cook. Behavior of bollworm (Lepidoptera: Noctuidae) larvae on genetically engineered cotton. *Journal of Economic Entomology* 95: 763-769 (2002).
 48. A.P. Gutierrez, L. Ponti, H.R. Herren, J. Baumgärtner, and P.E. Kenmore. Deconstructing Indian cotton: weather, yields, and suicides. *Environmental Sciences Europe* 27(12): 1-17 (2015).
 49. L. Lu, J. Luo, S. Zhang, Q. Yu, L. Ma, X. Liu, C. Wang, X. Ma, Y. Ma, and J. Cui. Efficiency of cotton bollworm (*Helicoverpa armigera* Hübner) control of different Bt cotton varieties in North China. *Journal of Cotton Research* 1: Article No. 4 (2018).
 50. C.N.K. Rajapakse, and G.H. Walter. Polyphagy and primary host plants: oviposition preference versus larval performance in the lepidopteran pest *Helicoverpa armigera*. *Arthropod-Plant Interactions* 1: 17-26 (2007).
 51. M.Ye, J. Beach, J.W. Martin, and A. Senthilselvan. Occupational pesticide exposures and respiratory health. *International Journal of Environmental Research and Public Health* 10: 6442-6471 (2013).
 52. Y. Carrière, J.A. Fabrick, and B.E. Tabashnik. Can Pyramids and Seed Mixtures Delay Resistance to Bt Crops? *Trends in Biotechnology* 34: 291-302 (2016).
 53. D. Singh, J. Gill, R. Gumber, R. Singh, and S. Singh. Yield and fibre quality associated with cotton leaf curl disease of Bt-cotton in Punjab. *Journal of Environmental Biology* 34: 113-116 (2013).
 54. M.U. Rahman, A.Q. Khan, Z. Rahmat, M.A. Iqbal, and Y. Zafar. Genetics and Genomics of Cotton Leaf Curl Disease, Its Viral Causal Agents and Whitefly Vector: A Way Forward to Sustain Cotton Fiber Security. *Frontiers Plant Sciences* 8: 1157 (2017).

55. M.N. Sattar, A. Kvarnheden, M. Saeed, and R.W. Briddon. Cotton leaf curl disease – an emerging threat to cotton production worldwide. *Journal of General Virology* 94: 695-710 (2013).
56. H.M. Nateshan, V. Muniyappa, M.M. Swanson, and B.D. Harrison. Host range, vector relations and serological relationships of cotton leaf curl virus from southern India. *Annals of Applied Biology* 128: 233-244 (1996).
57. R.C. Reddy, V. Muniyappa, J. Colvin, and S. Seal. A new begomovirus isolated from *Gossypiumbarbadense* in southern India. *Plant Pathology* 54(4): 570-570 (2005)
58. T. Hussain, and T. Mahmood. A note on leaf curl disease of cotton. *The Pakistan Cotton* 32: 248-251 (1988).
59. T. Hussain, M. Tahir, and T. Mahmood. Cotton leaf curl virus. *Pakistan Journal of Phytopathology* 3: 57-61 (1991).
60. K.K. Biswas, U.K. Bhattacharyya, S. Palchoudhury, N. Balram, A. Kumar, R. Arora, S.K. Sain, P. Kumar, R.K. Khetarpal, A. Sanyal, and P.K. Mandal. Dominance of recombinant cotton leaf curl Multan-Rajasthan virus associated with cotton leaf curl disease outbreak in northwest India. *PLoS One* 15: e0231886 (2020).
61. B. Nawaz, M. Naem, T.A. Malik, G. Muhae-Ud-Din, Q. Ahmad, and S. Sattar. A Review about Cotton Leaf Curl Viral Disease and Its Control Strategies in Pakistan. *International Journal of Innovative Approaches in Agricultural Research* 3: 132-147 (2019).
62. L. Amrao, S. Akhter, M.N. Tahir, I. Amin, R.W. Briddon and S. Mansoor. Cotton leaf curl disease in Sindh province of Pakistan is associated with recombinant begomovirus components. *Virus Research* 153: 161-165 (2010).
63. M. Ahmad, M.I. Arif, Z. Ahmad, and I. Denholm. Cotton whitefly (*Bemisia tabaci*) resistance to organophosphate and pyrethroid insecticides in Pakistan. *Pest Management Science* 58: 203-208 (2002).
64. W. Nazeer, S. Ahmad, K. Mahmood, A. Tipu, A. Mahmood, and B. Zhou. Introgression of genes for cotton leaf curl virus resistance and increased fiber strength from *Gossypium stocksii* into upland cotton (*G. hirsutum*). *Genetics and Molecular Research* 13: 1133-1143 (2014).
65. M.Ahmad, and G.E. Battese. A Probit Analysis of the Incidence of the Cotton Leaf Curl Virus in Punjab, Pakistan. *The Pakistan Development Review* 36: 155-169 (1997).
66. R. Peshin, B.S. Hansra, K. Singh, R. Nanda, R. Sharma, S. Yangsdon and R. Kumar. Long-term impact of Bt cotton: An empirical evidence from North India. *Journal of Cleaner Production* 312: 127575 (2021).
67. J.L. Hofs, A.S. Schoeman, and J. Pierre. Diversity and abundance of flower-visiting insects in Bt and non-Bt cotton fields of Maputaland (KwaZulu Natal Province, South Africa). *International Journal of Tropical Insect Science* 28: 211 (2008).
68. X. Men, F. Ge, C.A. Edwards, and E.N. Yardim. The influence of pesticide applications on *Helicoverpa armigera* Hübner and sucking pests in transgenic Bt cotton and non-transgenic cotton in China. *Crop Protection* 24: 319-324 (2005).
69. N.C. Naveen, R. Chaubey, D. Kumar, K.B. Rebijith, R. Rajagopal, B. Subrahmanyam, and S. Subramanian. Insecticide resistance status in the whitefly, *Bemisia tabaci* genetic groups Asia-I, Asia-II-1 and Asia-II-7 on the Indian subcontinent. *Scientific Reports* 7: 4063 (2017).
70. C. Sun, J. Xu, Q. Zhang, H. Feng, F. Wang, and R. Song. 2022. Effect of transgenic Bt cotton on population of cotton pests and their natural enemies in Xinjiang. *Chinese Journal of Biological Control* 18: 106-110 (2022).
71. M. Aslam, M. Razaq, N.A. Saeed, and F. Ahmad. Comparative resistance of different cotton varieties against bollworm complex. *International Journal of Agriculture and Biology* 6: 39-41 (2004).
72. G. Murtaza, M. Ramzan, U. Naem-Ullah, M.A. Qayyum, A. Nawaz, U.R. Azmi, and M. Ali. Population Dynamics of Cotton Jassid (*Amrassica Biguttula*) in Relation to Weather Parameters in Multan. *Acta Scientific Agriculture* 3: 212-215 (2019).
73. S.N. Harde, A.G. Mitkari, S.V. Sonune, and L.V. Shinde. Seasonal Incidence of Major Sucking Insect Pest in Bt Cotton and Its Correlation with Weather Factors in Jalna District (MS), India. *International Journal of Agriculture & Environmental Science* 5: 59-65 (2018).
74. M.R. Shahid, J. Farooq, A. Mahmood, F. Ilahi, M. Riaz, A. Shakeel, I.V. Petrescu-Mag, and A. Farooq. Seasonal occurrence of sucking insect pest in cotton ecosystem of Punjab, Pakistan. *Advances in Agriculture and Botany* 4: 26-30 (2012).
75. M.M. Zafar, A. Razzaq, M.A. Farooq, A. Rehman, H. Firdous, A. Shakeel, H. Mo, and M. Ren. Insect resistance management in *Bacillus thuringiensis* cotton by MGPS (multiple genes pyramiding and silencing). *Journal of Cotton Research* 3: 33 (2020).
76. F. Huang, D.A. Andow, and L.L. Buschman. Success of the high-dose/refuge resistance management strategy after 15 years of Bt crop use in North America. *Entomologia Experimentalis et Applicata* 140: 1-16 (2011).
77. Y. Chen, Y. Li, Y. Chen, E.H.M.A. Abidallha, D. Hu, Y. Li, X. Zhang, and D. Chen. Planting density

- and leaf-square regulation affected square size and number contributing to altered insecticidal protein content in Bt cotton. *Field Crops Research* 205: 14-22 (2017).
78. Y. Chen, Y. Li, M. Zhou, Q. Rui, Z. Cai, X. Zhang, Y. Chen, and D. Chen. Nitrogen (N) Application Gradually Enhances Boll Development and Decreases Boll Shell Insecticidal Protein Content in N-Deficient Cotton. *Frontiers Plant Sciences* 9: 51-51 (2018).
 79. L. Wang, Y. Ma, P. Wan, K. Liu, Y. Xiao, J. Wang, S. Cong, D. Xu, K. Wu, J.A. Fabrick, X. Li, and B.E. Tabashnik. Resistance to *Bacillus thuringiensis* linked with a cadherin transmembrane mutation affecting cellular trafficking in pink bollworm from China. *Insect Biochemistry and Molecular Biology* 94: 28-35 (2018).
 80. C. Luo, C.M. Jones, G. Devine, F. Zhang, I. Denholm, and K. Gorman. Insecticide resistance in *Bemisia tabaci* biotype Q (Hemiptera: Aleyrodidae) from China. *Crop Protection* 29: 429-434 (2010).
 81. A. Sethi, and V. K. Dilawari. Spectrum of Insecticide Resistance in Whitefly from Upland Cotton in Indian Subcontinent. *Journal of Entomology* 5: 138-147 (2008).
 82. T.C. Sparks, and R. Nauen. IRAC: Mode of action classification and insecticide resistance management. *Pesticide Biochemistry and Physiology* 121: 122-128 (2015).
 83. C. Erdogan, G. D. Moores, M.O. Gurkan, K.J. Gorman, and I. Denholm. Insecticide resistance and biotype status of populations of the tobacco whitefly *Bemisia tabaci* (Hemiptera: Aleyrodidae) from Turkey. *Crop Protection* 27: 600-605 (2008).
 84. E. Fernández, C. Grávalos, P.J. Haro, D. Cifuentes, and P. Bielza. Insecticide resistance status of *Bemisia tabaci* Q-biotype in south-eastern Spain. *Pest Management Science* 65: 885-891 (2009).
 85. M.Z. Khalid, S. Ahmed, I. Al-Ashkar, A. El Sabagh, L. Liu, and G. Zhong. Evaluation of Resistance Development in *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) in Cotton against Different Insecticides. *Insects* 12: 996 (2021).
 86. M. Khan, H. Mahmood, and C. Damalas. Pesticide use and risk perceptions among farmers in the cotton belt of Punjab, Pakistan. *Crop Protection* 67: 184-190 (2015).
 87. Government of Pakistan, *Economic Survey of Pakistan, Finance Division Islamabad, Pakistan* (2006). <https://www.finance.gov.pk/survey/chapters/02-Agriculture.PDF>.
 88. S. Rashid, W. Rashid, R.X.S. Tulcan, and H. Huang. Use, exposure, and environmental impacts of pesticides in Pakistan: a critical review. *Environmental Science and Pollution Research* 29(29): 43675-43689 (2022).
 89. M.I. Tariq. Leaching and degradation of cotton pesticides on different soil series of cotton growing areas of Punjab, Pakistan in Lysimeters. Ph. D. Thesis, *University of the Punjab, Lahore, Pakistan* (2005).
 90. M.I. Tariq, S. Afzal, I. Hussain, and N. Sultana. Pesticides exposure in Pakistan: a review. *Environment International* 33: 1107-1122 (2007).
 91. M.I. Tariq, S. Afzal, and I. Hussain. Pesticides in shallow groundwater of Bahawalnagar, Muzafargarh, D.G. Khan and Rajan Pur districts of Punjab, Pakistan. *Environment International* 30: 471-479 (2004).
 92. M. Ahmad, I. Arif, and M. Ahmad. Occurrence of insecticide resistance in field populations of *Spodoptera litura* (Lepidoptera: Noctuidae) in Pakistan. *Crop Protection* 26: 809-817 (2007).
 93. A. Tahir, F. Khan, and A. Khan. Effect of constant flow valves on performance of pesticide sprayers. *International Journal of Agriculture and Biology* 5: 49-52 (2003).
 94. C.A. Damalas, and I.G. Eleftherohorinos. Pesticide exposure, safety issues, and risk assessment indicators. *International Journal of Environmental Research and Public Health* 8: 1402-1419 (2011).
 95. D. Nuyttens, K. Baetens, M. De Schamphelleire, and B. Sonck. Effect of nozzle type, size and pressure on spray droplet characteristics. *Biosystems Engineering* 97: 333-345 (2007).
 96. J. Ferguson, A. Hewitt, and C. O'Donnell. Pressure, droplet size classification, and nozzle arrangement effect on coverage and droplet number density using air-inclusion dual fan nozzles for pesticide applications. *Crop Protection* 89: 231-238 (2016).
 97. D. Mada, and A. Medugu. Study on environmental impact of pesticides application with agricultural sprayers in southern Adamawa state- Nigeria. *IOSR Journal of Engineering* 4: 05-11 (2014).
 98. J. Wu, L. Ge, F. Liu, Q. Song, and D. Stanley. Pesticide-Induced Planthopper Population Resurgence in Rice Cropping Systems. *Annual Review of Entomology* 65: 409-429 (2020).
 99. M.D. Gogi, A.H. Syed, B. Atta, M. Sufyan, M.J. Arif, M. Arshad, and O.E. Liburd. Efficacy of biorational insecticides against *Bemisia tabaci* (Genn.) and their selectivity for its parasitoid *Encarsia formosa* Gahan on Bt cotton. *Scientific Reports* 11(1): 2101-2101 (2021).
 100. A. Nadeem, H.M. Tahir, A.A. Khan, A. Idrees, M.F. Shahzad, Z.A. Qadir, and J. Li. Response of natural enemies toward selective chemical insecticides; used for the integrated management of insect pests in cotton field plots. *Agriculture* 12(9): 1341-1341 (2022).
 101. S. Ejaz, W. Akram, C.W. Lim, J.J. Lee, and I. Hussain. Endocrine disrupting pesticides: a leading

- cause of cancer among rural people in Pakistan. *Experimental Oncology* 26: 98-105 (2004).
102. D. Shepard, M. Agnew, M. Fidanza, J. Kaminski, and L. Dant. Selecting nozzles for fungicide spray applications. *Golf Course Management* 74: 83-88 (2006).
 103. R. Klein, L. Schulze, and C. Ogg. Factors affecting spray drift of pesticides. *Crops Soils* 41: 19-23 (2008).
 104. M.S. Sharifzadeh, G. Abdollahzadeh, C.A. Damalas, and R. Rezaei. Farmers' criteria for pesticide selection and use in the pest control process. *Agriculture* 8: 24 (2018).
 105. T.J. Peters, A. Thostenson, J.F. Nowatzki, V.L. Hofman and J.A. Wilson. Selecting spray nozzles to reduce particle drift. *NDSU Extension Service, North Dakota State University* (2017). <https://www.sbreb.org/wp-content/uploads/2018/05/Selecting-Spray-Nozzles.pdf>.
 106. C.N. Kesavachandran, M. Fareed, M.K. Pathak, V. Bihari, N. Mathur, and A.K. Srivastava. Adverse Health Effects of Pesticides in Agrarian Populations of Developing Countries. *Reviews of Environmental Contamination and Toxicology* 200: 33-52 (2009).
 107. R. Chen, J. Huang, and F. Qiao. Farmers' knowledge on pest management and pesticide use in Bt cotton production in China. *China Economic Review* 27: 15-24 (2013).
 108. B.T. Mengistie, A.P. Mol, and P. Oosterveer. Pesticide use practices among smallholder vegetable farmers in Ethiopian Central Rift Valley. *Environment, Development and Sustainability* 19: 301-324 (2017).
 109. M.S. Allahyari, C.A. Damalas, and M. Ebadattalab. Farmers' technical knowledge about integrated pest management (IPM) in olive production. *Agriculture* 7(12): 101 (2017).
 110. C.E. Togbé, R. Haagsma, A.K. Aoudji, and S.D. Vodouhê. Effect of participatory research on farmers' knowledge and practice of ipm: the case of cotton in Benin. *The Journal of Agricultural Education and Extension* 21(5): 421-440 (2015).
 111. E.E. Lekei, A.V. Ngowi, and L. London. Farmers' knowledge, practices and injuries associated with pesticide exposure in rural farming villages in Tanzania. *BMC Public Health* 14: 1-13 (2014).
 112. A.V.F. Ngowi, T.J. Mbise, A.S. Ijani, L. London, and O.C. Ajayi. Smallholder vegetable farmers in Northern Tanzania: Pesticides use practices, perceptions, cost and health effects. *Crop Protection* 26: 1617-1624 (2007).
 113. E. Karamidehkordi, and A. Hashemi. Farmers' knowledge of IPM: A case study in the Zanjan Province in Iran. In: *Innovation and Sustainable Development in Agriculture and Food Symposium, ISDA, Montpellier, France. 10p. fffhal-00510402* (2010). <https://sid.ir/paper/601554/en>.
 114. M.Mubushar, F.O. Aldosari, M.B. Baig, B.M. Alotaibi, and A.Q. Khan. Assessment of farmers on their knowledge regarding pesticide usage and biosafety. *Saudi Journal of Biological Sciences* 26: 1903-1910 (2019).
 115. M.A. Khan, and M. Iqbal. Sustainable Cotton Production through Skill Development among Farmers: Evidence from Khairpur District of Sindh, Pakistan. *The Pakistan Development Review* 44: 695-716 (2005).
 116. F. Rehman, S. Muhammad, I. Ashraf, K.M. Ch, and T. Ruby. Effect of farmers' socioeconomic characteristics on access to agricultural information: Empirical evidence from Pakistan. *The Journal of Animal and Plant Sciences* 23(1): 324-329 (2013).
 117. S. Ashraf, G.A. Khan, S. Ali, S. Ahmed, and M. Iftikhar. Perceived effectiveness of information sources regarding improved practices among citrus growers in Punjab, Pakistan. *Pakistan Journal of Agricultural Sciences* 52: 861-866 (2015).
 118. O.Akinagbe, C. Attamah, and E. Igbokwe. Sources of Information on Climate Change among Crop Farmers in Enugu North Agricultural Zone, Nigeria. *International Journal of Research in Agriculture and Forestry* 2(11): 27-33 (2015).
 119. A. Abdullah. Analysis of mealybug incidence on the cotton crop using ADSS-OLAP (Online Analytical Processing) tool. *Computers and Electronics in Agriculture* 69: 59-72 (2009).
 120. G.H. Mallah, A.K. Keerio, A.R. Soomro, and A.W. Soomro. Population dynamics of predatory insects and biological control of cotton pests in Pakistan. *Journal of Biological Sciences* 1: 245-248 (2001).
 121. G.Abbas, M.J. Arif, M. Ashfaq, M. Aslam, and S. Saeed. Host plants, distribution and overwintering of cotton mealybug (*Phenacoccus Solenopsis*; Hemiptera: Pseudococcidae). *International Journal of Agriculture and Biology* 12: 421-425 (2010).
 122. A. Kamble, N. Danawale, and R. Kumar. Integrated weed management in Bt cotton. *Indian Journal of Weed Science* 49: 405-408 (2017).
 123. A. Mohyuddin, and M. Ambreen. From Faith Healer to a Medical Doctor: Creating Biomedical Hegemony. *Open Journal of Applied Sciences* 4: 56-67 (2014).
 124. J. Deguine, P. Ferron, and D. Russell. Sustainable pest management for cotton production. *A review. Agronomy for Sustainable Development* 28: 114-137 (2011).
 125. V.T. Sundaramurthy, and R.T. Gahukar. Integrated Management of Cotton Insect Pests in India. *Outlook on Agriculture* 27(4): 261-269 (1998).

126. K. Kranthi, and D. Russell. Changing trends in cotton pest management. Integrated Pest Management. In: Integrated Pest Management: Innovation-Development Process. R. Peshin, and A.K. Dhawan (Eds.). *Springer, Dordrecht* pp. 499-541 (2009).
127. M. Barzman, P. Bàrberi, A.N.E. Birch, P. Boonekamp, S. Dachbrodt-Saaydeh, B. Graf, and M. Sattin. Eight principles of Integrated Pest Management. *Agronomy for Sustainable Development* 35: 1199-1215 (2015).
128. M. Tudi, H.D. Ruan, L. Wang, J. Lyu, R. Sadler, D. Connell, C. Chu, and D.T. Phung. Agriculture Development, Pesticide Application and Its Impact on the Environment. *International Journal of Environmental Research and Public Health* 18: 1112 (2021).
129. J. Sherman, and D.H. Gent. Concepts of Sustainability, Motivations for Pest Management Approaches, and Implications for Communicating Change. *Plant Disease* 98: 1024-1035 (2014).
130. P.C. Matteson. Insect Pest Management in Tropical Asian Irrigated Rice. *Annual Review of Entomology* 45: 549-574 (2000).
131. S. Toepfer, T. Zhang, B. Wang, Y. Qiao, H. Peng, H. Luo, X. Wan, R. Gu, Y. Zhang, H. Ji, and M. Wan. Sustainable Pest Management through Improved Advice in Agricultural Extension. *Sustainability* 12: 6767 (2020).
132. Z. Korani. Application of Teaching Methods, Promoting Integrated Pest Management on the Farm School in Order to Achieve Sustainable Agriculture. *Procedia Social and Behavioral Sciences* 47: 2187-2191 (2012).